

# **Effect of catechin on lipid peroxidation and vital parameters of Holstein bull sperm after freeze-thawing process**

Sepehri-moghaddam, H.<sup>1</sup>; Emadi, M.<sup>1</sup> and Vafa, T.S.<sup>1</sup>

Received: 15.11.2017

Accepted: 12.06.2018

## **Abstract**

The aim of this study was to determine the antioxidant properties of catechin on lipid peroxidation and vital parameters of Holstein bull sperm after the freeze-thawing process. In this experimental study, semen samples were collected from four mature Holstein bull, twice a week using an artificial vagina. Ejaculates were pooled in order to eliminate the individual effects of bull. Semen samples were divided into four equal groups (8 reps). Zero (control), 20, 30 and 40 mg/ml of catechin with diluents based on egg yolk-citrate were added to the semen samples. Following cooling and equilibration stage of semen samples, the samples were stored in a nitrogen tank for a period of 30 days. After thawing procedure, the level of malondialdehyde in sperm samples were measured using the ELISA method. Also, membrane integrity, motility and viability of sperm were also examined. Results showed, membrane integrity, motility and viability of sperm samples treated with a concentration of 20, 30 and 40 mg/ml catechin in dose-dependent manner significantly increased and level of malondialdehyde dose-dependent manner significantly decreased, compared to the control groups. Therefore, the use of catechin in bull semen diluent can improve sperm vital parameters and decreases lipid peroxidation of sperm after the freeze-thawing process.

**Key words:** Catechin, Lipid Peroxidation, Bull, Sperm

---

1- Assistant Professor, Department of Animal Science, Faculty of Agriculture, Payam-e-Noor University, Tehran, Iran

**Corresponding Author:** Sepehri-moghaddam, H., E-mail: he.sepehrimoghadam@pnu.ac.ir

## References

- Abshenas, J.; Babaei, H.; Zare, M.H.; Allahbakhshi, A. and Sharififar, F. (2011). The effects of green tea (*Camellia sinensis*) extract on mouse semen quality after scrotal heat stress. Veterinary Research Forum, 2(4): 242-247.
- Amini-Pour, H.; Tahmasbi, A.M. and Naserain, A.A. (2013). The influence of vitamin E on semen characteristics of ghezel rams in during cooling and frozen process. European Journal of Zoological Research, 2(5): 94-99.
- Ashrafi, I.; Kohram, H. and Tayefi-Nasrabadi, H. (2013). Antioxidant effects of bovine serum albumin on kinetics, microscopic and oxidative characters of cryopreserved bull spermatozoa. Spanish Journal of Agricultural Research, 11(3): 695-701.
- Beheshti, R.; Yousefi Asl, M. and Ghiasi, J. (2012). Effects of addition of glutamine to semen extenders on microscopic factors of buffalo bulls. Journal of Comprehensive Photobiology, 9(38): 727-732.
- Chatterjee, S. and Gagnon C. (2001). Production of reactive oxygen species by spermatozoa undergoing cooling, freezing, and thawing. Molecular Reproduction and Development, 59(4): 451-458.
- Choi, S.J.; Park, S.Y.; Park, J.S.; Park, S.K. and Jung, M.Y. (2016). Contents and compositions of policosanols in green tea (*Camellia sinensis*) leaves. Food Chemistry, 204: 94-101.
- Daghikhia, H.; Shahbaz Zadeh, R. and Ashrafi, I. (2015). Antioxidant effect of Macrantha Satureja extraction on microscopic and biochemical parameters of bull sperm after freeze -thawing process. Pajouhesh and Sazandegi, 28(108): 101-112. (In Persian)
- De Graaf, S.P.; Evans, G.; Gillan, L.; Guerra, M.M.; Maxwell, W.M. and O'Brien, J.K. (2007). The influence of antioxidant, cholesterol and seminal plasma on the in vitro quality of sorted and non-sorted ram spermatozoa. Theriogenology, 67(2): 217-227.
- Dolati Doorbashi, P.; Moghaddam, G.; Daghikhia, H.; Taghizadeh, A. and Rafat S.A. (2015). The effect of adding different raffinose concentrations in the diluents in semen cryopreservation of different breeds of ram at the reproductive season. Animal Science Researches (Agricultural Sciences), 25(2): 109-132.
- Farhadi, F.; Towhidi, A. and Shakeri, M. (2015). The effect of adding different levels of zinc sulfate to semen diluent on quality of frozen-thawed sperm in bull. Iranian Journal of Animal Science, 45(4): 335-342.
- Farhadi, R.; Daghikhia, H.; Hosseinkhani, A.; Ghasemi Panahi, B.; Dehghan, G. and Ashrafi, I. (2016). Effect of Origanum vulgare ethanol extract on quality parameters and malondialdehyde concentration of cryopreserved Holstein bull sperm. Iranian Journal of Applied Animal Science, 6(4): 783-789.
- Gadkari, P.V. and Balaraman, M. (2015). Catechins: Sources, extraction and encapsulation: A review. Food and Bioproducts Processing, 93: 122-138.
- Gil, J.; Lundheim, N.; Soderquist, L. and Rodriuez-Martinez, H. (2003). Influence of extender, temperature, and addition of glycerol on post-thaw sperm parameters in ram semen. Theriogenology, 59(5-6): 1241-1255.
- Glasauer, A. and Chandel, N.S. (2014). Targeting antioxidants for cancer therapy. Biochemical Pharmacology, 92(1): 90-101.
- Ishii, T.; Mori, T.; Tanaka, T.; Mizuno, D.; Yamaji, R.; Kumazawa, S. et al. (2008). Covalent modification of proteins by green tea polyphenol (-) - epigallocatechin - 3 - gallate through autoxidation. Free Radical Biology & Medicine, 45(10): 1384-1394.
- Lambert, J.D. and Elias, R.J. (2010). The antioxidant and pro-oxidant activities of green tea polyphenols: a role in cancer prevention. Archives of Biochemistry and Biophysics, 501(1): 65-72.
- Liu, T.; Gao, J.; Zhou, N.; Mo, M.; Wang, X.; Zhang, X. et al. (2016). The effect of two cryopreservation methods on human sperm DNA damage. Cryobiology, 72(3): 210-215.
- Lucio, C.F.; Regazzi, F.M.; Silva, L.C.G.; Angriman, D.S.R.; Nichi, M. and Vannucchi C.I. (2016). Oxidative stress at different stages of two-step semen cryopreservation procedures in dogs. Theriogenology, 85(9): 1568-1575.
- Malek-Mohammadi, R.; Roghani, M. and Salami, M. (2015). The effect of aqueous extracts of *Melissa officinalis* on the oxidative stress indices in the midbrain tissue. Feyz, 19(1): 8-14.

- Malo, C.; Gil, L.; Cano, R.; Martínez, F. and Galé, I. (2011). Antioxidant effect of rosemary (*Rosmarinus officinalis*) on boar epididymal spermatozoa during cryopreservation. *Theriogenology*, 75(9): 1735-1741.
- Mehdipour, M.; Daghighe-Kia, H.; Najafi, A.; Vaseghi Dodaran, H. and García-Álvarez, O. (2016). Effect of green tea (*Camellia sinensis*) extract and pre-freezing equilibration time on the post-thawing quality of ram semen cryopreserved in a soybean lecithin-based extender. *Cryobiology*, 73(3): 297-303.
- Mocé, E. and Vicente, J.S. (2009). Rabbit sperm cryopreservation: A review. *Animal Reproduction Science*, 110(1-2): 1-24.
- Mohammadian, T.; Khodaei Motlagh, M. and Zare Shahneh, A. (2016). The effect of using different levels of royal jelly in semen extender on some quality and quantity parameters of Mahabadi goat semen. *Iranian Journal of Animal Science*, 46(4): 457-463.
- Mostek, A.; Dietrich, M.A.; Ślowińska, M. and Ciereszko, A. (2017). Cryopreservation of bull semen is associated with carbonylation of sperm proteins. *Theriogenology*, 92(1): 95-102.
- Oberoi, B.; Kumar, S. and Talwar, P. (2014). Study of human sperm motility post cryopreservation. *Medical Journal Armed Forces India*, 70(4): 349-353.
- Oliveira, P.F.; Tomás, G.D.; Dias, T.R.; Martins, A.D.; Rato, L.; Alves, M.G. et al. (2015). White tea consumption restores sperm quality in prediabetic rats preventing testicular oxidative damage. *Reproductive BioMedicine Online*, 31(4): 544-556.
- Purdy, P.H. (2006). A review on goat sperm cryopreservation. *Small Ruminant Research*, 63(3): 215-225.
- Purdy, P.H.; Ericsson, S.A.; Dodson, R.E.; Sternes, K.L. and Garner D.L. (2004). Effects of the flavonoids, silibinin and catechin, on the motility of extended cooled caprine sperm. *Small Ruminant Research*, 55(1-3): 239-243.
- Sadeghipanah, H.; Naijian, H.R. and Masoudi, R. (2015). Evaluation of antioxidant effects of butylated hydroxytoluene in the soybean lecithin based extender on frozen-thawed semen quality in Markhoz goat. *Iranian Veterinary Journal*, 11(2): 77-126. (In Persian)
- Seifi Jamadi, A.; Zareh, A.; Kohram, H.; Akbari, A.; Zamen, M. and Vakhideh, A. (2016). The potential of catalase as an enzymatic antioxidant to improve freezability of Turkmen stallions sperm. *Iranian Journal of Animal Science*, 42(2): 215-222.
- Shahbazzadeh, R.; Daghighe-Kia, H.; Moghaddam, G.; Dehghan, G.; Hosseinkhani, A. and Ashrafi, I. (2015). Effect of different levels of Satureja sahendica alcoholic extract on the quality of freeze-thawed Holstein bull spermatozoa. *Animal Science Researches (Agricultural Sciences)*, 25(1): 13-24.
- Shariatzadeh, M.A. and Mohammadi, M. (2015). Protective role of green tea (*Camellia sinensis*) hydroalcoholic extract on sperm parameters and testicular tissue in NMRI mice exposed to sodium arsenite. *Journal of Birjand University of Medical Sciences*, 21(4): 432-443.
- Spadiene, A.; Savickiene, N.; Ivanauskas, L.; Jakstas, V.; Skesters, A.; Silova, A. et al. (2014). Antioxidant effects of *Camellia sinensis* L. extract in patients with type 2 diabetes. *Journal of Food and Drug Analysis*, 22(4): 505-11.
- Swami, D.S.; Kumar, P.; Malik, R.K.; Saini, M.; Kumar, D. and Jan, M.H. (2017). The cryoprotective effect of iodixanol in buffalo semen cryopreservation. *Animal Reproduction Science*, 179: 20-26.
- Vilela, C.G.; Marquez, J.M.; Graham, J.K. and Barfield, J.P. (2017). Cryopreservation of bison epididymal sperm: A strategy for improving post-thaw quality when collecting sperm in field conditions. *Theriogenology*, 89: 155-161.
- Yue, H.X.; Li, P.; Jiang, M.; Lin, L. and Xu, K.H. (2005). Influence of cryopreservation with glycerol and freezing-thawing procedures on the motility of human sperm. *National journal of andrology*, 11(3): 204-206.