

Induction of estrus and fertility of Shal ewes using progesterone injections or intravaginal sponge followed by laparoscopic AI during non-breeding season

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Abstract

The objective of this study was to compare two estrus induction protocols in Shal ewes (Iranian native breed) during the non-breeding season using progesterone injections and an intravaginal sponge. Non pregnant ewes (n=180), considering their age and BCS were stratified randomly into two groups: progesterone injection (n=54; age: 33.0±3.21 months; BCS, 2.5±0.11) and vaginal sponge (n: 54; age: 37.5±3.58; BCS: 2.6±0.11). The progesterone injection group received four consecutive injections of progesterone (50 mg, SC), 3 days apart. Ewes in the sponge group received an intravaginal sponge (40 mg Flugestone acetate) for 14 days. An intramuscular injection of eCG (300 IU) was given concurrent with sponge withdrawal or 48 hrs after the last progesterone injection. Twelve hours after eCG, estrus was detected by rams equipped with harnesses. Laparoscopic AI was performed 50-56 hrs after eCG injection by fresh semen (100×10⁶ sperm in 0.2 ml per uterine horn). Pregnancy was diagnosed by rectal ultrasonography, 35 days after insemination. The incidence of estrus (74.1%) was greater in the sponge group than injection group (38.9%). There were no differences between the two groups in terms of time to estrus and time to AI after eCG injection. There was no differences between the two groups in the interval from estrus to AI, fertility, prolificacy, and fecundity. Fertility was greater in ewes that were inseminated within 24 hrs after estrous detection in the progesterone injected group (76.2%) compared to the sponge group (42.5%). In summary, due to the similarity in fecundity, as the final indicator of reproductive performance, and less cost, availability and no intervention of reproductive tract, progesterone injection method could be advised for inducing estrus in ewes during the non-breeding season.

Key word: Estrus induction, Progesterone injection, Sponge, Non-breeding season, Shal ewes

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References

- Abecia, J. A., Forcada, F., & González-Bulnes, A. (2011). Pharmaceutical control of reproduction in sheep and goats. *Veterinary Clinics: Food Animal Practice*, 27(1): 67-79.
- Abecia, J. A., Forcada, F., & González-Bulnes, A. (2012). Hormonal control of reproduction in small ruminants. *Animal Reproduction Science*, 130(3-4): 173-179.
- Ali, A. (2007). Effect of time of eCG administration on follicular response and reproductive performance of FGA-treated Ossimi ewes. *Small Ruminant Research*, 72(1): 33-37.
- Boland, M. P., Crosby, T. F., & Gordon, I. (1981). Effect of mating management and PMSG dose on lambing outcome in ewes bred in late anoestrus. *The Journal of Agricultural Science*, 97(2): 445-447.
- Bragança, J. F. M., Drissen, R. O., Machado, S. A., Bennemann, P. E., & da Rocha, R. X. (2019). Efficacy of the re-utilization of an ear implant impregnated with progesterone in estrus synchronization response and pregnancy in sheep. *Tropical Animal Health and Production*, 51(6): 1763-1765.
- Dutt, R. H. (1953). Induction of estrus and ovulation in anestrual ewes by use of progesterone and pregnant mare serum. *Journal of Animal Science*, 12(3): 515-523.
- Evans, G., & Maxwell, W. C. (1987). *Salamons' Artificial Insemination of Sheep and Goats* (No. Ed. 2). Butterworth-Heinemann.
- Farrag, B., El-Hawy, A. S., El-Bassiony, M. F., El-Rayes, M. A. H., & Shedeed, H. A. (2018). Influence of short- and long-term administration of Melengestrol acetate on estrus activity and reproductive performance of nulliparous Barki ewes. *International Journal of Environment, Agriculture and Biotechnology*, 3(4): 264403.
- Gharibi, S., Niasari-Naslaji, A., Poursasan, N., & Moosavi-Movahedi, A. A. (2014). Replacement of Salamon with Shotor diluent and egg yolk with low density lipoprotein for chilled storage of ram semen. *Iranian Journal of Veterinary Research*, 15: 279-284.
- Godfrey, R. W., Gray, M. L., & Collins, J. R. (1997). A comparison of two methods of oestrous synchronisation of hair sheep in the tropics. *Animal Reproduction Science*, 47(1-2): 99-106.
- Greyling, J. P. C., & Brink, W. C. J. (1987). Synchronization of oestrus in sheep: The use of controlled internal drug release (CIDR) dispensers. *South African Journal of Animal Science*, 17(3): 128-132.
- Greyling, J. P. C., Kotze, W. F., Taylor, G. J., Hagendijk, W. J., & Cloete, F. (1994). Synchronization of oestrus in sheep: use of different doses of progestagen outside the normal breeding season. *South African Journal of Animal Science*, 24(1): 33-37.
- Hashemi, M., Safdarian, M., & Kafi, M. (2006). Estrous response to synchronization of estrus using different progesterone treatments outside the natural breeding season in ewes. *Small Ruminant Research*, 65(3): 279-283.
- Hill, J. R., Thompson, J. A., & Perkins, N. R. (1998). Factors affecting pregnancy rates following laparoscopic insemination of 28,447 Merino ewes under commercial conditions: a survey. *Theriogenology*, 49(4): 697-709.
- Koyuncu, M., & Ozis Alticekic, S. (2010). Effects of progestagen and PMSG on estrous synchronization and fertility in Kivircik ewes during natural breeding season. *Asian-Australasian Journal of Animal Sciences*, 23(3): 308-311.
- López-García, S., Sánchez-Torres, M. T., Cordero-Mora, J. L., Figueroa-Velasco, J. L., Martínez-Aispuro, J. A., García-Cué, J. L., & Cárdenas-León, M. (2021). Estrous synchronization in sheep with reused progesterone devices and eCG. *Revista Brasileira de Zootecnia*, 50.
- Manes, J., Hozbor, F., Alberio, R., & Ungerfeld, R. (2014). Intravaginal placebo sponges affect negatively the conception rate in sheep. *Small Ruminant Research*, 120(1): 108-111.
- Martinez-Ros, P., Lozano, M., Hernandez, F., Tirado, A., Rios-Abellan, A., López-Mendoza, M. C., & Gonzalez-Bulnes, A. (2018). Intravaginal device-type and treatment-length for ovine estrus synchronization modify vaginal mucus and microbiota and affect fertility. *Animals*, 8(12): 226.
- Moakhar, H. K., Kohram, H., Shahneh, A. Z., & Saberifar, T. (2012). Ovarian response and pregnancy rate following different doses of eCG treatment in Chall ewes. *Small Ruminant Research*, 102(1): 63-67.
- Najafi, G., Cedden, F., & Maleki, S. (2014). The determination of plasma progesterone, estradiol-17 β hormone levels in Ghezel sheep treated with CIDR and various doses of PMSG during the breeding season. *Bulletin of Environment, Pharmacology and Life Sciences*, 3(3): 118-122.

- Niasari-Naslaji, A., & Soukhtezari, A. (2005). Comparison between three estrus synchronization programs using progestagens during the breeding season in the ewe. *Pajouhesh Sazandegi*, 65: 86-91.
- Noakes, D. E., Parkinson, T. J., & England, G. C. (2018). Puberty and seasonality. In: *Veterinary Reproduction and Obstetrics*. (10th ed). Elsevier Health Sciences. P. 54-62.
- Petrović, M. P., Caro Petrović, V., Ružić-Muslić, D., Maksimović, N., Ilić, Z. Z., Milošević, B., & Stojković, J. (2012). Some important factors affecting fertility in sheep. *Biotechnology in Animal Husbandry*, 28(3): 517-528.
- Rastegarnia, A., & Heydari, H. (2010). Oestrus synchronization in ewes with Fluorogestone acetate out of breeding season. *Journal of Large Animal Clinical Science Research*, 4(11): 21-30.
- Safdarian, M., Kafi, M., & Hashemi, M. (2006). Reproductive performance of Karakul ewes following different oestrous synchronisation treatments outside the natural breeding season. *South African Journal of Animal Science*, 36(4): 229-234.
- Sareminejad, P., Tabatabaei Vakili, S., Mamouei, M., Mirzadeh, Kh., & Boujarpour, M. (2015). Status of estrus and blood serum estrogen and progesterone in Arabic ewes synchronized with CIDR in non-breeding season. *Journal of Animal Science and Research*, 25(1): 151-161.
- SAS, Statistical Analysis System. (2012). User's Guide, version 9.4. SAS Institute, Cary, NC.
- Seidi-Samani, H., Niasari-Naslaji, A., Vojgani, M., Ganjkanlou, M., Alijani, A., Baninajjar, M. (2023). Synchronization of estrus using progesterone injections followed by hMG in ewes. *Veterinary Research Forum*, 14 (3): 145-151.
- Skliarov, P., Pérez, C., Petrusha, V., Fedorenko, S., & Bilyi, D. (2021). Induction and synchronization of oestrus in sheep and goats. *Journal of Central European Agriculture*, 22(1): 39-53.
- Swelum, A. A. A., Alowaimer, A. N., & Abouheif, M. A. (2015). Use of fluorogestone acetate sponges or controlled internal drug release for estrus synchronization in ewes: Effects of hormonal profiles and reproductive performance. *Theriogenology*, 84(4): 498-503.
- Youngquist, R.S., & Threlfall, W.R. (2007). *Current Therapy in Large Animal Theriogenology*. (2 nd ed). Saunders Elsevier. Philadelphia, USA. p. 642-704.