

# Effect of oral administration of different concentrations *Lactobacillus acidophilus* on growth performance and digestive enzyme rainbow trout fish (*Oncorhynchus mykiss*) in the face of lead toxicity in the diet

Mohammadian, T.<sup>1</sup>; Mohiseni, M.<sup>2</sup>; Ahmadi, B.<sup>3</sup> and Zeai Nejad, S.<sup>4</sup>

Received: 17.04.2017

Accepted: 01.11.2017

## Abstract

The present study was conducted to evaluate the effects of probiotic *Lactobacillus acidophilus* on the growth performance and digestive enzymes activity of juveniles *Oncorhynchus mykiss* after lead poisoning. 375 fish about  $16\pm 3.8$  g of weight were selected and after being physically examined and ensuring their health were divided randomly in to 5 groups including 3 groups which were fed by healthy food containing  $5\times 10^6$ ,  $5\times 10^7$  and  $5\times 10^8$  CFU/g *Lactobacillus acidophilus* for 45 days And also the fourth group (negative control or uncontrolled control) were fed and stored throughout the experiment with food without any additives And the fifth group (positive control or control group with lead), at first were fed without any additive to the diet for 45 days, and then, until the end of the experiment period, with three probiotic groups for 21 days were fed with a diet containing 500  $\mu\text{g}/\text{kg}$  lead nitrate. Intestine samples after anesthesia were collected on days 0, 45, 52, 59 and 66 for digestive enzymes were examined. Results showed that In the Treatment 1 and 2, Specific Growth Ratio, Daily Weight Growth and Relative Growth Rate, after 45 days beginning of experiment, improved considerable that compared to control group had significantly difference ( $P<0.05$ ). Activity of digestive enzyme in trial treatment (1, 2, and 3) after 45 days, were increased significantly compared to control group ( $P<0.05$ ). But after challenge with lead, trypsin and  $\alpha$ -Amylase decrease significantly in groups 1 and 2 but in upper than the other groups. The lipase enzyme and alkaline phosphatase activity in all the probiotic groups after exposure to lead to a significant decrease but more than control group was challenged to lead. According to obtained results, it might be concluded that the feeding of rainbow trout by  $5\times 10^7$  CFU / gr *Lactobacillus acidophilus* isolated from the intestinal *Tor gryp* could be used as a probiotic supplement and can have a positive impact on improving growth performance and digestive enzymes and in addition are preventing of heavy metal poisoning which exist in diet.

**Key word:** Probiotic, *Lactobacillus acidophilus*, *Oncorhynchus mykiss*, Enzyme, Lead

---

1- Assistant Professor, Department of Clinical Sciences, Faculty of Veterinary Medicine, Shahid Chamran University of Ahvaz, Ahvaz, Iran

2- Assistant Professor, Department of Fisheries, Faculty of Natural Resources, Behbahan Khatam Alanbia University of Technology, Behbahan, Iran

3- MSc. Graduated of Fisheries Faculty of Natural Resources, Behbahan Khatam Alanbia University of Technology, Behbahan, Iran

4- Associated Professor, Department of Fisheries, Faculty of Natural Resources, Behbahan Khatam Alanbia University of Technology, Behbahan, Iran

**Corresponding Author:** Mohammadian, T., E-mail: takavar\_m2002@yahoo.com

## References

- Al-Dohail, M.A.; Hashim, R. and Aliyu-Paiko, M. (2009). Effects of the probiotic, *Lactobacillus acidophilus*, on the growth performance, haematology parameters and immunoglobulin concentration in African catfish (*Clarias gariepinus*, Burchell 1822) fingerlings. *Aquaculture Research*, 40 (11): 1642-1652.
- Aly, S.M.; Abdel-Galil, A.Y.; Abdel-Aziz, Gh.A. and Mohamed, M.F. (2008). Studies on *Bacillus subtilis* and *Lactobacillus acidophilus*, as potential probiotics, on the immune response and resistance of *Tilapia nilotica* (*Oreochromis niloticus*) to challenge infections. *Fish and Shellfish Immunology*, 25 (1-2): 128-136.
- Axelsson, L. (1998). Lactic acid bacteria: classification and physiology. In: Salminen S and Von Wright A (Eds) *Lactic Acid Bacteria: Microbiology and Functional Aspects*, 2<sup>nd</sup> ed. New York. Marcel Dekker, Pp: 1-72.
- Balcazar, J.L.; Blas, I.; Ruiz-Zarzuola, I.; Cunningham, D.; Vandrell, D. and Muzquiz, J.L. (2006). Review: The role of probiotics in aquaculture. *Veterinary Microbiology*, 114 (3-4): 173-186.
- Bernfeld, P. (1955). Amylases  $\alpha$  and  $\beta$ . In: *Methods in enzymology*. Colowick, P and Kaplan, N.O. (Eds). 1<sup>st</sup> ed. New York. Academic press, Pp: 149-157.
- Cha, J.; Rahimnejad, S.; Yang, S.; Kim, K. and Lee, K. (2013). Evaluations of *Bacillus* spp. as dietary additives on growth performance, innate immunity and disease resistance of olive flounder (*Paralichthys olivaceus*) against *Streptococcus iniae* and as water additives. *Aquaculture*, 402-403: 50-57.
- Cahu, C.L.; Zambonino Infante, J.L.; Quazuguel, P. and Le Gall, M.M. (1999). Protein hydrolysate vs. fish meal in compound diets for 10-day old sea bass *Dicentrarchus labrax* larvae. *Aquaculture*, 171 (1-2): 109-111.
- Crespo, S.; Nonnotte, G.; Colin, D.A.; Leray, C.; Nonnotte, L. and Aubre, A. (1986). Morphological and functional alterations induced in trout intestine by dietary cadmium and lead. *Journal of Fish Biology*, 28 (1): 69-80.
- Elsanhoty, R.M.; Al-Turki, I.A. and Ramadan, M.F. (2016). Application of lactic acid bacteria in removing heavy metals and aflatoxin B1 from contaminated water. *Water Science and Technology*, 74 (3): 625-38.
- Erlanger, B.F.; Kokowsky, N. and Cohen, W. (1961). The preparation and properties of two new chromogenic substrates of trypsin. *Archives of Biochemistry and Biophysics*, 95 (2): 271-278.
- Gerbino, E.; Carasi, P.; Tymczyszyn, E. and Gómez-Zavaglia, A. (2014). Removal of cadmium by *Lactobacillus kefir* as a protective tool against toxicity. *Journal of Dairy Research*, 81 (3): 280-287.
- Giri, S.S.; Sukumaran, V. and Oviya, M. (2013). Potential probiotic *Lactobacillus plantarum* VSG3 improves the growth, immunity, and disease resistance of tropical freshwater fish, *Labeo rohita*. *Fish and Shellfish Immunology*, 34 (2): 660-666.
- Hauville, M.R.; Zambonino, J.L.; Gordon, B.J.; Migaud, H. and Main, K.L. (2016). Effects of a mix of *Bacillus* sp. as a potential probiotic for Florida pompano, common snook and red drum larvae performances and digestive enzyme activities. *Aquaculture Nutrition*, 22(1): 51-60.
- Kinoshita, H.; Sohma, Y.; Ohtake, F.; Ishida, M.; Kawai, Y.; Kitazawa, H. et al. (2013). Biosorption of heavy metals by lactic acid bacteria and identification of mercury binding protein. *Research in Microbiology*, 164 (7): 701-709.
- Kuz'mina, V.; Shekovtsova, N. and Bolobonina, V. (2010). Activity dynamics of proteinases and glycosidases of fish chyme with exposure in fresh and brackish water. *The Biological Bulletin*, 37 (6): 605-611.
- Lee, J.; Cheng, H.; Damte, D.; Lee, S.; Kim, J.; Rhee, M. et al. (2013). Effects of dietary supplementation of *Lactobacillus pentosus* PL11 on the growth performance, immune and antioxidant systems of Japanese eel *Anguilla japonica* challenged with *Edwardsiella tarda*. *Fish and Shellfish Immunology*, 34 (3): 756-761.
- Mohammadian, T.; Alishahi, M.; Tabandeh, M.R.; Ghorbanpoor, M.; Gharibi, D.; Tollabi, M. and Rohanzade, S. (2016). Probiotic effects of *Lactobacillus plantarum* and *L. delbrueckii* ssp. *bulguricus* on some immune-related parameters in *Barbus grypus*. *Aquaculture International*, 24(1): 225-242.

- Mohammadian, T.; Alishahi, M.; Tabandeh, M.R.; Ghorbanpoor, M.; Gharibi, D.; Tollabi, M. and Rohanzade, S. (2017). Effect of *Lactobacillus plantarum* and *Lactobacillus delbrueckii* subsp. *bulgaricus* on growth performance, gut microbial flora and digestive enzymes activities in *Tor grypus* (Karaman, 1971). Iranian Journal of Fisheries Sciences, 16 (1): 296-317.
- Mrvčić, J.; Stanzer, D.; Šolić, E. and Stehlik-Tomas, V. (2012). Interaction of lactic acid bacteria with metal ions: opportunities for improving food safety and quality. World Journal of Microbiology and Biotechnology, 28 (9): 2771-2782.
- Panwichian, S.; Kantachote, D.; Wittayaweerarak, B. and Mallavarapu, M. (2011). Removal of heavy metals by exopolymeric substances produced by resistant purple nonsulfur bacteria isolated from contaminated shrimp ponds. Electronic Journal of Biotechnology, 14 (4): 2-2.
- Planas, M.; Vazquez, J.A.; Marques, J.; Peres-Lomba, R.; Gonzalez, M.P. and Murado, M. (2004). Enhancement of rotifer (*Brachionus plicatilis*) growth by using terrestrial lactic acid bacteria. Aquaculture, 240 (1-4): 313-329.
- Ramos, M.A.; Weber, B.; Gonçalves, J.F.; Santos, G.A.; Rema, P. and Ozório, R.O.A. (2013). Dietary probiotic supplementation modulated gut microbiota and improved growth of juvenile rainbow trout (*Oncorhynchus mykiss*). Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology, 166 (2): 302-307.
- Rayes, A.A.H. (2012). Field studies on the removal of lead, cadmium and copper by the use of probiotic lactic acid bacteria from the water for culturing marine tilapia *T. spilurus*. New York Science Journal, 5 (11): 74-82.
- Rungruangsak-Torrissen, K.; Rustad, A.; Sunde, J.; Eiane, S.A.; Jensen, H.B.; Opstvedt, J. et al. (2002). In vitro digestibility based on fish crude enzyme extract for prediction of feed quality in growth trials. Journal of the Science of Food and Agriculture, 82 (6): 644-654.
- Safari, R.; Adel, M.; Lazado, C.C.; Caipang, C.A.M. and Dadar, M. (2016). Host-derived probiotics *Enterococcus casseliflavus* improves resistance against *Streptococcus iniae* infection in rainbow trout (*Oncorhynchus mykiss*) via immunomodulation. Fish and Shellfish Immunology, 52: 198-205.
- Sastry, K.V. and Gupta, P.K. (1978). In vitro inhibition of digestive enzymes by heavy metals and their reversal by chelating agent: Part I. Mercuric chloride intoxication. Bulletin of Environmental Contamination and Toxicology, 20 (1): 729-735.
- Singh, A.L. and Sarma, P.N. (2010). Removal of arsenic (III) from waste water using *Lactobacillus acidophilus*. Bioremediation Journal, 14 (2): 92-97.
- Soleimani, N.; Hoseinifar, S.H.; Merrifield, D.L.; Barati, M. and Hassan Abadi, Z. (2012). Dietary supplementation of fructooligosaccharide (FOS) improves the innate immune response, stress resistance, digestive enzyme activities and growth performance of Caspian roach (*Rutilus rutilus*) fry. Fish and shellfish immunology, 32 (2): 316-21.
- Son, V.M.; Chang, C.C.; Wu, M.C.; Guu, Y.K.; Chiu, C.H. and Cheng, W. (2009). Dietary administration of the probiotic, *Lactobacillus plantarum*, enhanced the growth, innate immune responses, and disease resistance of the grouper *Epinephelus coioides*. Fish and Shellfish Immunology, 26 (5): 691-698.
- Sunde, J.; Taranger, G. and Rungruangsak-Torrissen, K. (2001). Digestive protease activities and free amino acids in white muscle as indicators for feed conversion efficiency and growth rate in Atlantic salmon (*Salmo salar* L.). Fish Physiology and Biochemistry, 25 (4): 335-345.
- Suzer, C.; Çoban, D.; Kamaci, H.O.; Saka, S.; Firat, K. and Otgucuoğlu, Ö. (2008). *Lactobacillus* spp. Bacteria as probiotics in gilthead sea bream (*Sparus aurata*, L.) larvae: effects on growth performance and digestive enzyme activities. Aquaculture, 280 (1-4): 140-5.
- Venkat, H.K.; Sahu, N.P. and Jain, K.K. (2004). Effect of feeding *Lactobacillus*-based probiotics on the gut microflora, growth and survival of postlarvae of *Macrobrachium rosenbergii* (de Man). Aquaculture Research, 35 (5): 501-507.
- Vijavabaskar, P. and Somasundaram, S. (2008). Isolation of bacteriocin producing lactic acid bacteria from fish gut and probiotic activity against common fresh water fish pathogen *Aeromonas hydrophila*. Biotechnology, 7 (1): 124-128.

Vine, N.G.; Leukes, W.D.; Kaiser, H.; Daya, S.; Baxter, J. and Hecht, T. (2004). Competition for attachment of aquaculture candidate probiotic and pathogenic bacteria on fish intestinal mucus. *Journal of Fish Diseases*, 27 (6): 319-326.

Worthington, C.C. (1991). *Worthington enzyme manual related Biochemical*. New Jersey. 3<sup>rd</sup> ed. Freehold, Pp: 212-215.