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## Molecular prevalence of *Anaplasma* species in Cow of Mazandaran province

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## **Abstract**

This study aimed to determinate the variety of *Anaplasma* species among cows of Mazandaran province. For this purpose, 105 blood samples randomly collected via the jugular vein from different parts of Mazandaran province. The extracted DNA from blood cells was amplified by *Anaplasma*-all primers, which amplify an approximately 1468bp DNA fragment from a region of the 16S rRNA gene from various members of the genus *Anaplasma*. 29 (27.6%) out of 105 cow blood samples were *Anaplasma* spp. positive by first PCR and nested PCR. All cow positive samples were analysed for the presence of *A. phagocytophilum*, *A.bovis*, and *A.centrale* (*Amori Strain*) and as a result, 22 of blood samples (21%) for *A.phagocytophilum*, 12 (11.4%) for *A.bovis* and 1(1%) for *A.centrale* were positive. The extracted DNA from positive *Anaplasma spp* samples were amplified by *Anaplasma marginale* specific primers, which amplify an approximately 866bp DNA fragment from a region of the *msp4* gene. Out of 105, blood samples, Five (4.8%) were positive for *Anaplasma marginale*. This study is the first molecular detection of *Anaplasma* species from cows in Mazandaran province. The results show that there is a significant difference between the percentage of infection with *Anaplasma phagocytophilum*, in different seasons of the year and the livestock type. Also, the percentage of infection with *Anaplasma bovis*, among the variables studied; there is a significant difference between the seasons of the year and the type of livestock.

Key words: Molecular Prevalence, Anaplasma species, Cow, Mazandaran

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## Refrences

- Aubry, P., & Geale, D. (2011). A review of bovine anaplasmosis. *Transboundary and emerging diseases*, 58(1), 1-30.
- Baráková, I., Derdáková, M., Carpi ,G., Rossom ,F., Collini ,M., Tagliapietra ,V. et al. (2014). Genetic and ecologic variability among Anaplasma phagocytophilum strains, northern Italy. *Emerging Infectious Diseases*. 2014; 20:1082–4.
- Bashiribod, H., Kazemi ,K., Eslami ,G., Bigdeli, S., Bandehpour, M., Rahbarian ,N., et al.(2004). First molecular detection of Anaplasma phagocytophilum in Ixodes ricinus ticks in Iran. *Journal of medical sciences*. 2004; 4(4): 282-286.
- Bock, R.E., de Vos, A.J., Kingston, T.G., Carter ,P.D. (2003). Assessment of a low virulence Australian isolate of Anaplasma marginale for pathogenicity, immunogenicity and transmissibility by Boophilus microplus. *Veterinary Parasitology*, 118, 121–131.
- Carelli, G., Decaro, N., Lorusso, E., Paradies, P., Elia, G., Martella, V., . . . Ceci, L. (2008). First report of bovine anaplasmosis caused by anaplasma centrale in europe. *Annals of the New York Academy of Sciences*, 1149(1), 107-110
- Dumler, J. S., Madigan, J. E., Pusterla, N., & Bakken, J. S. (2007). Ehrlichioses in humans: Epidemiology, clinical presentation, diagnosis, and treatment. *Clinical infectious diseases*, 45(Supplement\_1), S45-S51
- Dumler, J.S., Barbet, A.F., Bekker, C.P., Dasch, G.A., Palmer, G.H., Ray, S.C., . . . Rurangirwa, F.R. (2001). Reorganization of genera in the families Rickettsiaceae and Anaplasmataceae in the order Rickettsiales: unification of some species of Ehrlichia with Anaplasma, Cowdria with Ehrlichia and Ehrlichia with Neorickettsia, descriptions of six new species combinations and designation of Ehrlichia equi and 'HGE agent' as subjective synonyms of Ehrlichia phagocytophilia. *International Journal of Systematic and Evolutionary Microbiology* 51: 2145-2165.
- Engvall, E.O., & Egenvall, A. (2002). Granulocytic ehrlichiosis in Swedish dogs and horses. *International Journal of Medical Microbiology* 291: 100-103.
- Jonsson, N. N., and S. W.J. Reid. (2000). Global climate change and vector borne diseases. *The Veterinary Journal*. 160, 87–89
- Kocan, K.M., de la, F. J., Blouin, E.F., Coetzee, J.F., Ewing, S.A. (2010). The natural history of Anaplasma marginale. *Veterinary Parasitology*. 2010; 167:95–107.
- Kocan, K.M., Blouin, E.F., Barbet, A.F. (2008). Anaplasmosis control. Past, present, and future. *Annals of the New York Academy of Sciences*. 2000; 916:501–9.
- Kocan, K. M., De la ,F. J., Guglielmone, A. A., & Meléndez, R. D. (2003). Antigens and alternatives for control of anaplasma marginale infection in cattle. *Clinical microbiology reviews*, 16(4), 698-71
- Ladbury, G. A., Stuen, S., Thomas, R., Bown, K. J., Woldehiwet, Z., Granquist, E. G., . . . Birtles, R. J. (2008). Dynamic transmission of numerous anaplasma phagocytophilum genotypes among lambs in an infected sheep flock in an area of anaplasmosis endemicity. *Journal of Clinical Microbiology*, 46(5), 1686-1691
- Liu, Z., Ma, M., Wang, Z., Wang, J., Peng, Y., Li, Y., . . . Yin, H. (2012). Molecular survey and genetic identification of anaplasma species in goats from central and southern china. *Applied and environmental microbiology*, 78(2), 464-470
- Masuzawa, T., Uchishima, Y., Fukui, T., Okamoto, Y., Pan, M. J., Kadosaka, T., & Takada, N. (2014). Detection of anaplasma phagocytophilum and anaplasma bovis in small wild mammals from taichung and kinmen island, taiwan. *Japanese journal of infectious disease* .67, 111-114, 2014
- Matei, I. A., Ionică, A. M., D'Amico, G., Corduneanu, A., Daskalaki, A. A., Lefkaditis, M., & Mihalca, A. D. (2017). Altitude-dependent prevalence of canine granulocytic anaplasmosis in romania. *Vector-Borne and Zoonotic Diseases*.17 (2):147-151.
- M'ghirbi, Y., Yaïch, H., Ghorbel, A., & Bouattour, A. (2012). Anaplasma phagocytophilum in horses and ticks in tunisia. *Parasites & Vectors*, 5(1), 1-7
- Noaman, V. (2017). A review of anaplasmosis and the prevalence of anaplasma marginale in cattle in iran and the world. *Veterinary Researches & Biological Products*, 30(3), 2-15 (In Persian).

- Noaman, V. (2013). Report of anaplasma centrale (amori strain) in cattle in iran. *Pajouhesh-va-Sazandegi Veterinary Journal*, 98, 26-29. (In Persian).
- Noaman, V.,& Moradi, M. (2019). Molecular epidemiology and risk factors assessment of anaplasma spp. On dairy cattle in southwest of iran. *Acta Veterinaria Eurasia*, 46(1), 30-37
- Noaman, V., & Shayan, P. (2010). Molecular detection of anaplasma bovis in cattle from central part of iran. *Veterinary Research Forum* Vol: 1, No: 2, September, 2010, 117 - 122
- Noaman, V., Shayan, P., (2009). Molecular detection of Anaplasma phagocytophilum in carrier cattle of Iran first documented report. *Iranian Journal of Microbiology*. 1(2): 37-42
- Noaman, V., S. Arabzadeh and B. Kachooei. (2002). A study on Anaplasmosis in cattle of Falavarjan city, Isfahan province (1995-200). *Pajouhesh Va- Sazandegi* 51:10-12. (In Persian).
- Ooshiro, M., Zakimi, S., Matsukawa, Y., Katagiri, Y., Inokuma, H. (2008). Detection of Anaplasma bovis and Anaplasma phagocytophilum from cattle on Yonaguni Island, Okinawa, Japan. *Veterinary Parasitology* 154: 360–364
- Pace, J., & Wakeman, D. (2003). Determining the age of cattle by their teeth, university of florida, if as extension.
- Park1, J., Han, D. G., Ryu, J.H., Chae, J. B., Chae J,S.b., Yu, D.H., . . . Choi ,K. S. (2018). Molecular detection of Anaplasma bovis in Holstein cattle in the Republic of Korea. *Acta Veterinaria Scandinavica* (2018) 60:15
- Pazhoom, F., Ebrahimzade, E., Shayan, P., Nabian, S. (2016). Anaplasma spp. identification in hard ticks of Iran: First report of Anaplasmabovis in Haemaphysalis inermis. *Acarologia* 56(4): 497–504
- Rajput. Z.I., Hu Song-hua, A.G., Arijo, H., Habib and K Khalid, (2005). Comparative study of Anaplasma parasites in tick carrying buffaloes and cattle. *Journal of Zhejiang University-Scince A*, 6B: 1057-1062.
- Renneker, S., Abdo, J., Salih, D.E.A., Karagenc, T., Bilgic, H., Torina, A et al. (2013). Can Anaplasma ovis in small ruminants be neglected any longer? *Transboundary and Emerging Diseases*. 2013; 60:105–12.
- Rymaszewska, A. (2011). PCR For detection of tick-borne Anaplasma phagocytophilum pathogens: *a review. Veterinarni Medicina*. 2011; 56:529–36.
- Sarih, M., M'Ghirbi, Y., Bouattour, A., Gern, L., Baranton, G., Postic, D. (2005). Detection and identification of Ehrlichia spp. in ticks collected in Tunisia and Morocco. *Journal of Clinical Microbiology*. 2005; 43:1127–32.
- Stuen, S., Granquist, E.G., Silaghi, C. (2013). Anaplasma phagocytophilum a widespread multi-host pathogen with highly adaptive strategies. *Frontiers in Cellular and Infection Microbiology*. 2013; 3:1127–32.
- Stuen, S. (2007). Anaplasma phagocytophilum-the most widespread tick-borne infection in animals in europe. *Veterinary research communications*, 31(1), 79-84
- Stuen, S., Bergström, K., Petrovec, M., Van de Pol, I., & Schouls, L. M. (2003). Differences in clinical manifestations and hematological and serological responses after experimental infection with genetic variants of anaplasma phagocytophilum in sheep. *Clinical and Vaccine Immunology*, 10(4), 692-695
- Stuen, S., Bergstrom, K. (2001). Serological investigation of granulocytic Ehrlichia infection in sheep in Norway. *Acta Veterinaria Scandinavica*, 42:331-338
- Tana-Hernández, L., Navarrete-Arroyo, K., Ron-Román, J., Reyna-Bello, A., & Chávez-Larrea, M. A. (2017). Pcr-diagnosis of anaplasma marginale in cattle populations of ecuador and its molecular identification through sequencing of ribosomal 16s fragments. BMC *veterinary research*, 13(1), 1-7
- Teshale, S., Geysen, D., Ameni, G., Bogale, K., Dorny, P., & Berkvens, D. (2016). Molecular detection of anaplasma species in questing ticks (ixodids) in ethiopia. *Asian Pacific Journal of Tropical Disease*, 6(6), 449-452
- Torina, A., Caracappa, S. (2007). Anaplasmosis in cattle in Italy. *Veterinary Research Communications*, 31(Suppl.1): 73–78.
- Torina, A., Vicente, J., Alongi, A., Scimeca, S., Turlá, R., Nicosia, S., Di Marco, V., Caracappa, S. & De La Fuente, J. (2007). Observed prevalence of tick-borne pathogens in domestic animals in Sicily, Italy during 2003–2005. *Zoonoses and Public Health*. 2007; 54:8–15.

- Vahedi Noori, N. Abdi, G, M., Mohammad Nejad, K. Sh. (2016). Evaluation of the species diversity and abundance of hard ticks (Family: Ixodidae) parasite of cattle and sheep in Mazandaran province. *Veterinary Journal (Pajouhesh & Sazandegi)* No 106 pp: 58. (In Persian).
- Weisburg, W. G., Barns, S. M., and Pelletier, D. A., (1991). 16S ribosomal DNA amplification for phylogenetic study. *Journal of Bacteriology*. 1991; 173: 697-703.
- Yang, J., Li, Y., Liu, Z., Liu, J., Niu, Q., Ren, Q Yang, J., Li, Y., Liu, Z., Liu, J., Niu, Q., Ren, Q., Chen, Z., Guan, G., Luo, J. & Yin, H. (2015). Molecular detection and characterization of anaplasma spp. In sheep and cattle from xinjiang, northwest china. *Parasites & Vectors*, 8(1), 1-7.
- Yang, J., Liu, Z., Guan, G., Liu, Q., Li, Y., Chen, Z., Ma, M., Liu, A., Ren, Q., Luo, J. & Yin, H. (2013). Prevalence of anaplasma phagocytophilum in ruminants, rodents and ticks in gansu, north-western china. *Journal of medical microbiology*, 62(2), 254-258